

m FC Agar (NCM0149)

Intended Use

m FC Agar is used with rosolic acid for the detection and enumeration of fecal coliforms by membrane filtration in a laboratory setting. m-FC Agar is not intended for use in the diagnosis of disease or other conditions in humans.

Description

Geldreich et al. formulated a medium to enumerate fecal coliforms (FC) using the membrane filter (m) technique without prior enrichment.

Many standard method membrane filtration procedures recommend m-FC media for testing water. The American Public Health Association (APHA) specified m-FC media and incubation at $44.5 \pm 0.5^\circ\text{C}$ in the fecal coliform procedure and other tests. The Association of Official Analytical Chemists (AOAC) specifies m-FC Agar for detecting total coliforms and fecal coliforms in foods. The US Environmental Protection Agency specified using m-FC media in fecal coliform methods for testing water by the direct MF method or the delayed-incubation MF methods.

Typical Formulation

Enzymatic Digest of Casein	10.0 g/L
Enzymatic Digest of Animal Tissue	5.0 g/L
Yeast Extract	3.0 g/L
Sodium Chloride	5.0 g/L
Lactose	12.5 g/L
Bile Salts	1.5 g/L
Aniline Blue	0.1 g/L
Agar	15.0 g/L

Final pH: 7.4 ± 0.2 at 25°C

Formula may be adjusted and/or supplemented as required to meet performance specifications.

Supplement

1% Rosolic Acid, 10 mL

Precaution

Refer to SDS

Preparation

1. Suspend 52 g of the medium in 1 L of purified water containing 10 mL of 1% Rosolic Acid in 0.2 N NaOH.
2. If necessary, adjust pH to 7.4 with 1N HCl.
3. Heat with frequent agitation and boil for one minute to completely dissolve the medium.
4. Cool to $45 - 50^\circ\text{C}$ and pour plates.
5. DO NOT AUTOCLAVE.

Rosolic Acid: Dissolve 1 g in 100 mL of 0.2 N NaOH to prepare a 1% solution.

Quality Control Specifications

Dehydrated Appearance: Powder is homogeneous, free flowing, and pale to light grey-blue to greyish-beige.

Prepared Appearance: Prepared unsupplemented medium is dark blue and clear to slightly hazy. Prepared appearance with 1% Rosolic Acid is trace to slightly hazy and purple to reddish purple to cranberry red.

Expected Cultural Response: Cultural response incubated aerobically on m-FC Agar at $44.5 \pm 0.5^\circ\text{C}$ and examined for growth after 22 - 24 hours.

Microorganism	Approx. Inoculum (CFU)	Expected Results	
		Growth (with Rosolic Acid)	Reaction
<i>Enterobacter faecalis</i> ATCC® 19433	10 - 100	Completely Inhibited	---
<i>Escherichia coli</i> ATCC® 11775	10 - 100	Good	Dark blue colonies
<i>Escherichia coli</i> ATCC® 25922	10 - 100	Good	Dark blue colonies
<i>Salmonella typhimurium</i> ATCC® 14028	10 - 100	Good	Reddish-grey colonies
<i>Staphylococcus aureus</i> ATCC® 25923	1000	Completely Inhibited	---

The organisms listed are the minimum that should be used for quality control testing.

Test Procedure

1. Filter duplicate samples through separate membrane filters.
2. Transfer filters to surface of separate m-FC Agar plates.
3. Place each plate in a separate waterproof plastic bag. Submerge in waterbath at $44.5 \pm 0.5^\circ\text{C}$; incubate for 22 - 24 hours.

Results

Fecal coliforms will be various shades of dark blue. Non-fecal coliforms are grey to reddish-grey to cream-colored.

Expiration

Refer to expiration date stamped on the container. The dehydrated medium should be discarded if not free flowing, or if medium has changed from the original color. Expiry applies to medium in its intact container.

Limitations of the Procedure

1. Due to varying nutritional requirements, some strains may be encountered that grow poorly or fail to grow.
2. A few non-fecal coliform colonies may be observed on m-FC Agar due to the selective action of the elevated temperature and the addition of rosolic acid. It may be useful to elevate the temperature to $45 \pm 0.2^\circ\text{C}$ to eliminate *Klebsiella* strains from the fecal coliform group.

Storage

Store dehydrated culture media at $2-30^\circ\text{C}$ away from direct sunlight. Once opened and recapped, place container in a low humidity environment at the same storage temperature. Protect from moisture and light by keeping container tightly closed.

References

1. Geldreich, E. E., H. F. Clark, C. B. Huff, and L. C. Best. 1965. Fecal-coliform-organism medium for the membrane filter technique. *J. Am. Water Works Assoc.* 57:208-214.
2. Eaton, A. D., L. S. Clesceri, and A. E. Greenberg (eds.). 2017. *Standard methods for the examination of water and wastewater*, 23rd ed. American Public Health Association, Washington, D.C.
3. Cowman, S., and R. Kelsey. 1992. Bottled water, p. 1031-1036. *In* C. Vanderzant, and D. F. Splittstoesser (eds.). *Compendium of methods for the microbiological examination of foods*, 3rd ed. American Public Health Association, Washington, D.C.
4. Andrews, W. 2016. *Microbial methods*. In *Official methods of analysis of AOAC International*, 20th ed. AOAC International. Arlington, VA.

Technical Specification Sheet



5. Bordner, R., and J. Winter (eds.). 1978. Microbiological methods for monitoring the environment. EPA-600/8-78-017. Environmental Monitoring and Support Laboratory, Office of Research and Development, U. S. Environmental Protection Agency, Cincinnati, OH.
6. Environmental Protection Agency. 1992. Manual for the certification of laboratories analyzing drinking water. EPA-814B-92-002. Office of Ground Water and Technical Support Division, U. S. Environmental Protection Agency, Cincinnati, OH.

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