# LAB

## LAB195

# BCYE Legionella Isolation Medium

#### DESCRIPTION

BCYE (Buffered Charcoal Yeast Extract) Legionella Isolation Medium (LAB195) is a base medium used for the isolation of Legionella from clinical and environmental samples. This medium is based on the charcoal yeast extract formulation of Feeley *et al.*<sup>1&2</sup>. The performance of this medium is further enhanced by the additions of ACES (N-2-acetamido-2aminoethane - sulphonic acid) buffer and a-ketoglutarate as defined by Edelstein<sup>3</sup>. This medium is also detailed in internationally recoanized methodology<sup>4</sup> for the isolation of Legionella spp. from water.

Specimens or samples are often heavily contaminated with other bacteria and consequentially a range of selective supplements have been developed to aid isolation. Lab M provide the GVPC supplement (X195) which is most effective for the isolation of L. pneumophila. It is recommended that this supplement is used in conjunction with heat and acid sample treatments, to further reduce the growth of non-Legionella bacteria.

This product contains the ACES

BCYE Legionella Isolation Medium (LAB195)		
Formula	Amount/ Litre	
Yeast Extract Charcoal Ferric Pyrophosphate	10.00g 2.00g 0.25g	
ACES Buffer Potassium Carbonate	10.00g 2.28g	
Agar	14.00g	
Supplements		
GVPC Selective (X195)		
Glycine Vancomycin Polymyxin B Cycloheximide	3000mg 1mg 79200IU 80mg	
Growth Supplement (X196)		
L-Cysteine a-ketoglutarate	400mg 1000mg	
Presumptive ID	(X197)	
a-ketoglutarate	1000mg	
Add one vial sterilised medium		
рН 6.9 <u>+</u> 0.1		



buffer and ferric pyrophosphate in the base medium. This negates the need for complex freeze dried supplements. A complementary growth supplement is provided (X196) which contains the Lcysteine and a-ketoglutarate. In a-ketoalutarate addition, an supplement (X197) is also available for the preparation of confirmatory media for suspected Legionella colonies.

### PRINCIPLE OF ISOLATION

Water samples are concentrated either by membrane filtration or centrifugation (turbid samples may also be centrifuged). То reduce the growth of unwanted bacteria, separate portions of the concentrated sample may be subjected to heat and acid treatments. Treated and untreated portions then are inoculated onto Legionella selective media.

#### METHOD FOR RECONSTITUTION

Selective Isolation. Weigh 38.5 grams of powder and disperse in 1 litre of deionised water. Soak for 10 minutes, swirl to mix and sterilise by autoclaving at 110°C for 10 minutes. Cool to 47°C and aseptically add 2 vials of reconstituted growth supplement X196 and 2 vials of reconstituted



selective supplement X195. Mix well and pour into sterile Petri dishes.

*Maintenance.* Weigh 38.5 grams of powder and disperse in 1 litre of deionised water. Soak for 10 minutes, swirl to mix and sterilize by autoclaving at 110°C for 10 minutes. Cool to 47°C and aseptically add 2 vials of reconstituted growth supplement X196. Mix well and pour into Petri dishes.

Presumptive Identification. Weigh 38.5 grams of powder and disperse in 1 litre of deionised water. Soak for 10 minutes, swirl mix and sterilize to bv autoclaving at 110°C for 10 minutes. Cool to 47°C and aseptically add 2 vials of reconstituted growth supplement X197. Mix well and pour into Petri dishes.

### INOCULATION

The concentrated sample should be split into 3 portions. One portion is used without any further treatment, the other 2 portions should be treated, one with heat and the other with acid. *Heat Treatment*. Take 1ml of the concentrated sample and place in a water bath at 50°C for 30 minutes.

Acid Treatment. Take 1-10ml of the concentrated sample and centrifuge at 6000g for 10 minutes. Decant the supernatant to leave half the original volume. Vortex to re-suspend the pellet and make up to the original volume using an HCI-KCI buffer. Leave to stand for 5 minutes.

Inoculate the first plate of GVPC supplemented media with 0.1mL of the untreated portion and spread over the entire surface of the plate. Inoculate the second

plate of GVPC supplemented media in the same way with 0.1ml of the heat treated portion as soon as possible after removal from the water bath. Inoculate the third plate of GVPC supplemented media in the same way with 0.1mL of the acid treated portion immediately after acid treatment.

#### **INCUBATION**

Incubate at  $36 \pm 1^{\circ}$ C in a humid atmosphere under aerobic conditions for up to 10 days.

### INTERPRETATION

The plates should be examined for growth on days 3, 5, 7 and 10. Suspect colonies should be sub-cultured on to "maintenance" supplemented BCYE medium and "presumptive ID" supplemented BCYE medium, incubate as before. Isolates that fail to grow on the "presumptive ID" medium but grow on the maintenance medium and have typical morphology should be regarded as presumptive Legionella.

Presumptive isolates should be confirmed using a serological method, e.g. Microgen M45 Latex.

MINIMUM Q.C. ORGANISMS *Legionella* spp. - Growth

*Staphylococcus epidermidis* Growth

Escherichia coli - Inhibited

#### PACKAGING

BCYE	Legionella	Isolation
Medium:		

1 litre sample	LAB195-E38.5
500g	LAB195-A
2.5kg	LAB195-B2.5
5kg	LAB195-B5
10kg	LAB195-B10

*X195 GVPC Selective Supplement* - 10 vials, each sufficient for 500ml of media.

*X196 BCYE Growth Supplement* - 10 vials, each sufficient for 500ml of media.

*X197 BCYE Growth Supplement* (omitting L-Cysteine) - 10 vials, each sufficient for 500ml of media.

*LATEX KITS*: Product M45 . Lab M also distributes the Microscreen® Legionella Latex Kit. This provides reagents for the confirmation of *Legionella pneumophila* serogroups 1 and 2-15.

#### REFERENCES

1. Feeley, J.C., Gibson, R.J. *et al.* (1979). *Journal of Clinical Microbiology* **10**: 437-441

2. Pesculle, A.H., Feeley, J.C. *et al.* (1980). *Journal of Infectious Disease* **141**: 727-732

3. Edelstein, P.H. (1982). Journal of Clinical Microbiology 14: 298-303

4. International Standard. ISO 11731:1998(E). Water Quality – Detection & Enumeration of *Legionella*.